

# Highly sensitive targeted and non-targeted HPLC-MS analysis of PFAS

Per- and polyfluoroalkyl substances (PFAS) are persistent environmental pollutants known for their long lifetime and mobility. Their stability leads to accumulation in groundwater and soil, with proven harmful health effects. To mitigate environmental pollution, several PFAS compounds, such as perfluorooctanoic acid (PFOA), perfluorohexanesulfonic acid (PFHxS), and long-chain perfluoroalkyl carboxylic acids (C9-C14), are now regulated.

This technical note presents a method for identifying and quantifying PFAS using a YMC-Triart C18 HPLC column. This column, based on a robust hybrid silica particle, offers enhanced separation of isomers and improved analytical performance when coupled with mass spectrometry.





#### Table 1: Chromatographic conditions.

Column: YMC-Triart C18 (12 nm, 3 µm) 100 x 2.1 mm ID

Part No.: TA12S03-10Q1PTH

Eluent: A) water/methanol (95/5) + 2 mM ammonium acetate B) water/methanol (5/95) + 2 mM ammonium acetate

Gradient: 15-70 %B (0-2 min), 70-90 %B (2-5 min), 90-100 %B (5-10 min), 100 %B (10-15 min),

15 %B (15.1-22 min)

Flow rate: 0.3 mL/min Temperature:  $40 \,^{\circ}$ C Injection:  $2 \,\mu$ L

Detection: ESI-MS negative mode

Sample: PFAS standards (5, 10, 25, 50, 75, 100 ng/mL) from Fluka, Sigma, Dr. Ehrenstorfer, Apollo Scientific,

soil extract sample (Brilon-Scharfenberg)

#### Table 2: MS source parameters.

Gas Temp: 150 °C Gas Flow: 16 L/min 35 psig Nebulizer pressure: Sheath gas temperature: 380 °C 12 L/min Sheath gas flow: Fragmentor voltage: 380 V 3000 V Capillary voltage: Nozzle voltage: 300 V

Table 3: Substances included in the standard mixture.

Name	Abbreviation	Mass (Da)	m/z	Retention time (s)
Perfluorobutanoic acid	PFBA	213.9865	212.9792	291
Perfluoropentanoic acid	PFPeA	263.9833	262.9760	359
Perfluorohexanoic acid	PFHxA	313.9801	312.9728	398
Perfluoroheptanoic acid	PFHpA	363.9769	362.9696	432
Perfluorooctanoic acid	PF0A	413.9737	412.9664	469
Perfluorononanoic acid	PFNA	463.9705	462.9632	505
Perfluorodecanoic acid	PFDA	513.9673	512.9600	538
Perfluorobutanesulfonic acid	PFBS	299.9503	298.9430	366
Perfluorohexanesulfonic acid	PFHxS	399.9439	398.9366	435
Perfluorooctanesulfonic acid	PF0S	499.9375	498.9302	503
6:2 Fluorotelomer sulfonic acid	FTSA	427.9752	426.9679	467



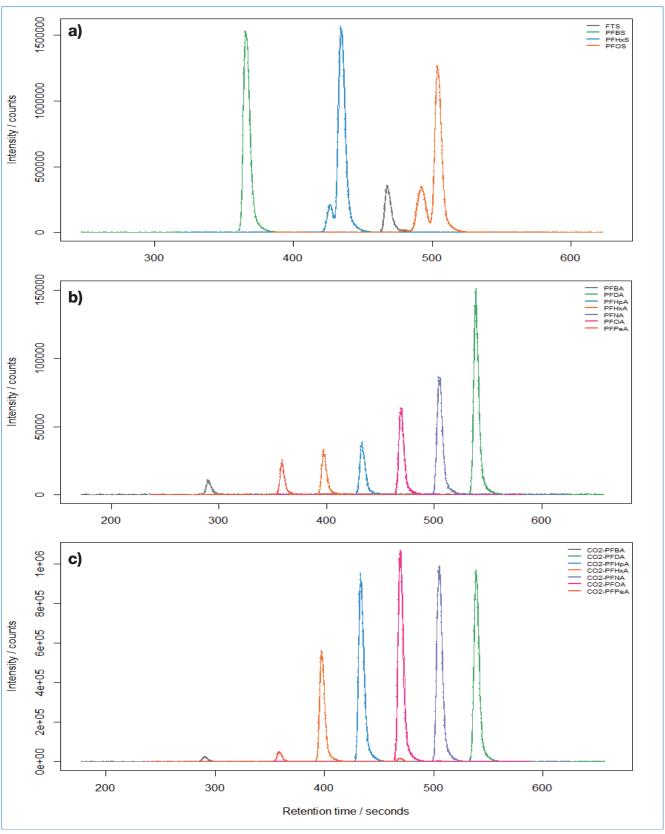


Figure 1: Extracted ion chromatograms of (a) perfluoroalkyl sulfonic acids (PFSAs), (b) perfluoroalkyl carboxylic acids (PFCAs) and (c) perfluoroalkyl carboxylic acids (PFCAs) with loss of  ${\rm CO_2}$  during ionisation obtained with +/-120 seconds and +/-15 ppm at a concentration of 75 ng/ml each.



PFAS standards (Table 3) were analysed at concentrations ranging from 5–100 ng/mL. The standards could be well resolved by the YMC-Triart column (Figure 1). Notably, partial separation of PFOS and PFHxS isomers within 10 seconds facilitated fragmentation pattern analysis for structural elucidation.

For perfluoroalkyl carboxylic acids (PFCAs), higher peak intensities were observed for longer carbon chains as

expected (Figure 1b). Additionally,  $\rm CO_2$ -loss ions exhibited remarkably higher intensities than the primary molecular ions (Figure 1c). Calibration curves (Figure 2) demonstrated a linear quantification range, and carry-over tests confirmed no residual analytes after injections of 100 ng/mL. These results highlight the YMC-Triart C18 column's suitability for robust PFAS analysis.

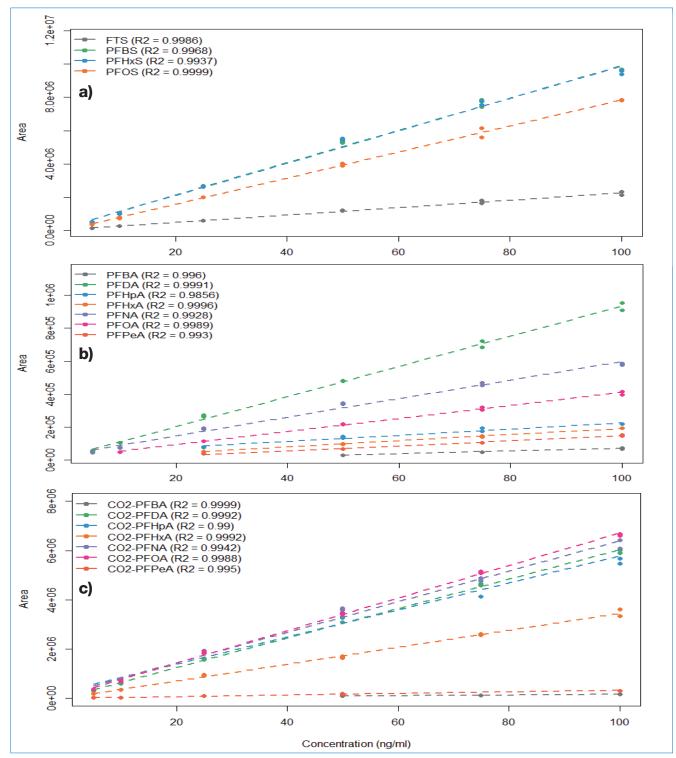


Figure 2: Calibration curves of (a) perfluoroalkyl sulfonic acids (PFSAs), (b) perfluorinated carboxylic acids (PFCAs) and (c) perfluorinated carboxylic acids (PFCAs) with loss of  $\mathrm{CO}_2$  during ionisation at concentrations of 5-100 ng/mL.



### Non-targeted analysis in a soil extract

To evaluate performance in complex matrices, a soil extract from a contaminated site (Brilon-Scharfenberg, Germany; from the scientific publication of Zweigle et al. [1]) was analysed. Total ion chromatograms (TICs) from triplicate measurements (Figure 3) revealed consistent profiles, despite the high matrix complexity. Table 4 lists pre-identified PFAS clusters detected in the analysis (originally identified in Zweigle et al. [1] and shared by courtesy).

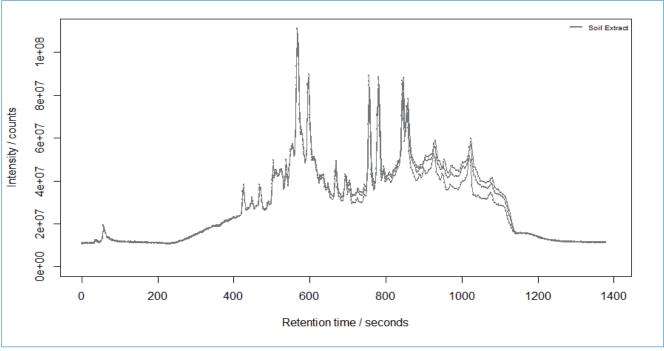


Figure 3: Total ion chromatograms (TIC) of the soil sample measured in triplicate.



Table 4: Substances identified in the soil extract by Zweigle et al. [1], using Kendrick mass defect analysis and matching of  $CF_2$ -distances in the fragmentation spectra of prioritised chromatographic peaks.

•	·	
m/z	Cluster Group	
448.93286		
498.92966		
548.92647		
598.92328	F(OF ) 00	
648.92008	F(CF <sub>2</sub> ) <sub>n</sub> SO <sub>3</sub>	
698.9168		
748.91369		
798.9105		
848.90731		
506.901728		
556.898528		
606.895328		
656.892128	SF <sub>5</sub> (CF <sub>2</sub> ) <sub>0</sub> SO <sub>3</sub>	
706.888928		
756.885728		
806.882528		
460.932838		
510.929638		
560.926438		
610.923238		
660.920038	CF(CF <sub>2</sub> ) <sub>0</sub> SO <sub>3</sub>	
710.916838		
760.913638		
810.910438		
860.907238		
	448.93286 498.92966 548.92647 598.92328 648.92008 698.9168 748.91369 798.9105 848.90731 506.901728 556.898528 606.895328 656.892128 706.888928 756.885728 806.882528 460.932838 510.929638 560.926438 610.923238 660.920038 710.916838 760.913638 810.910438	

Formula	m/z	Cluster Group	
CF <sub>3</sub> O(CF <sub>2</sub> ) <sub>8</sub> SO <sub>3</sub>	564.921359	CF <sub>3</sub> O(CF <sub>2</sub> ) <sub>n</sub> SO <sub>3</sub>	
CF <sub>3</sub> O(CF <sub>2</sub> ) <sub>9</sub> SO <sub>3</sub>	614.918159		
CF <sub>3</sub> O(CF <sub>2</sub> ) <sub>10</sub> SO <sub>3</sub>	664.914959		
CF <sub>3</sub> O(CF <sub>2</sub> ) <sub>11</sub> SO <sub>3</sub>	714.911759		
CF <sub>3</sub> O(CF <sub>2</sub> ) <sub>12</sub> SO <sub>3</sub>	764.908559		
CF <sub>3</sub> O(CF <sub>2</sub> ) <sub>13</sub> SO <sub>3</sub>	814.905359		
CF <sub>3</sub> O(CF <sub>2</sub> ) <sub>14</sub> SO <sub>3</sub>	864.902159		
CF <sub>3</sub> O(CF <sub>2</sub> ) <sub>15</sub> SO <sub>3</sub>	914.898959		
CF <sub>3</sub> OC <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>6</sub> SO <sub>3</sub>	526.924553	CF <sub>3</sub> OC <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>n</sub> SO <sub>3</sub>	
CF <sub>3</sub> OC <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>7</sub> SO <sub>3</sub>	576.921353		
CF <sub>3</sub> OC <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>8</sub> SO <sub>3</sub>	626.918153		
CF <sub>3</sub> OC <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>9</sub> SO <sub>3</sub>	676.914953		
CF <sub>3</sub> OC <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>10</sub> SO <sub>3</sub>	726.911753		
CF <sub>3</sub> OC <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>11</sub> SO <sub>3</sub>	776.908553		
CF <sub>3</sub> OC <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>12</sub> SO <sub>3</sub>	826.905353		
CF <sub>3</sub> OC <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>13</sub> SO <sub>3</sub>	876.902153		
FC <sub>2</sub> F <sub>2</sub> C <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>4</sub> SO <sub>3</sub>	422.936031	FC <sub>2</sub> F <sub>2</sub> C <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>n</sub> SO <sub>3</sub>	
FC <sub>2</sub> F <sub>2</sub> C <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>6</sub> SO <sub>3</sub>	522.929631		
FC <sub>2</sub> F <sub>2</sub> C <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>7</sub> SO <sub>3</sub>	572.926431		
FC <sub>2</sub> F <sub>2</sub> C <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>8</sub> SO <sub>3</sub>	622.923231		
FC <sub>2</sub> F <sub>2</sub> C <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>9</sub> SO <sub>3</sub>	672.920031		
FC <sub>2</sub> F <sub>2</sub> C <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>10</sub> SO <sub>3</sub>	722.916831		
FC <sub>2</sub> F <sub>2</sub> C <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>11</sub> SO <sub>3</sub>	772.913631		
FC <sub>2</sub> F <sub>2</sub> C <sub>2</sub> F <sub>2</sub> (CF <sub>2</sub> ) <sub>12</sub> SO <sub>3</sub>	822.910431		



### **Improved separation of isomers**

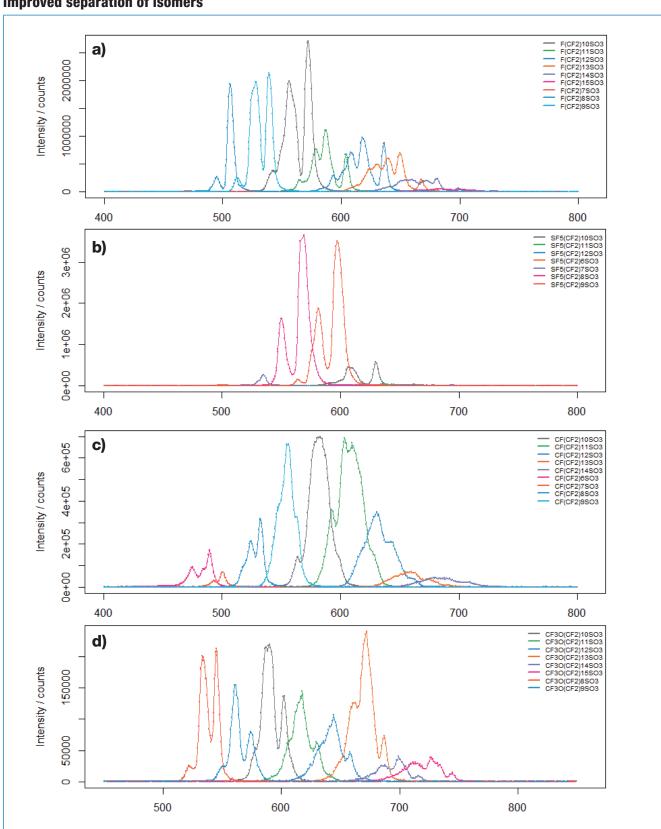


Figure 4: Extracted ion chromatograms of (a) PFAS group  $F(CF_2)_nSO_3$ , (b) PFAS group  $SF_5(CF_2)_nSO_3$ , (c) PFAS group  $CF(CF_2)_nSO_3$ , (d) PFAS group  $CF_3O(CF_2)_nSO_3$ , (e) PFAS group  $CF_3O(CF_2)_nSO_3$ , (e) PFAS group  $CF_3O(CF_2)_nSO_3$ .



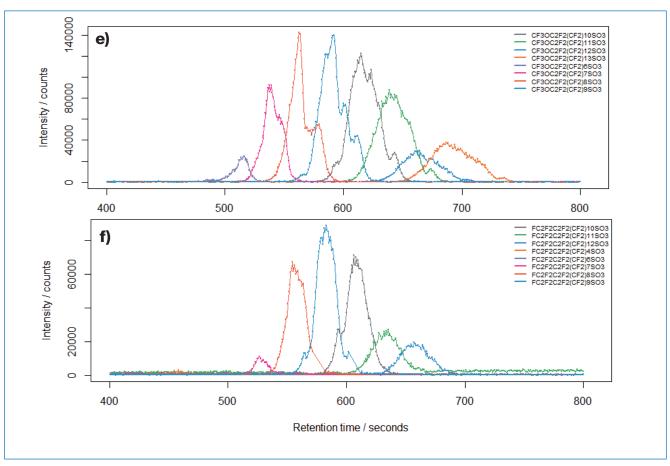


Figure 4: Extracted ion chromatograms of (a) PFAS group  $F(CF_2)_nSO_3$ , (b) PFAS group  $SF_5(CF_2)_nSO_3$ , (c) PFAS group  $CF_3(CF_2)_nSO_3$ , (d) PFAS group  $CF_3(CF_2)_nSO_3$ , (e) PFAS group  $CF_3(CF_2)_nSO_3$  and (f) PFAS group  $FC_2F_2C_2F_2(CF_2)_nSO_3$  continued.

The YMC-Triart C18 column achieved effective separation of PFAS (Figure 4), including improved partial resolution of isomers compared to Zweigle et al. [1]. For instance,  $SF_5(CF_2)_9SO_3$  isomers were separated (Figure 5), enabling the acquisition of distinct fragmentation spectra (Figure 6). Ion traces at 575 s and 600 s revealed differences in

fragmentation patterns, indicating structural variations. Identifiable fragments are listed in Table 5. The larger fragments in longer retention indicate an exclusively linear isomer. Specifically, the shorter retention time and the smaller fragments suggest a branched isomer or a differently positioned  $SF_5$ -group.

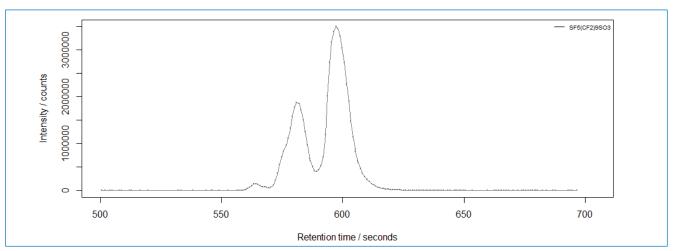


Figure 5: Ion trace correspondent to  $SF_5(CF_2)_9SO_3^-$  with m/z 656.8927 +/-20 ppm.



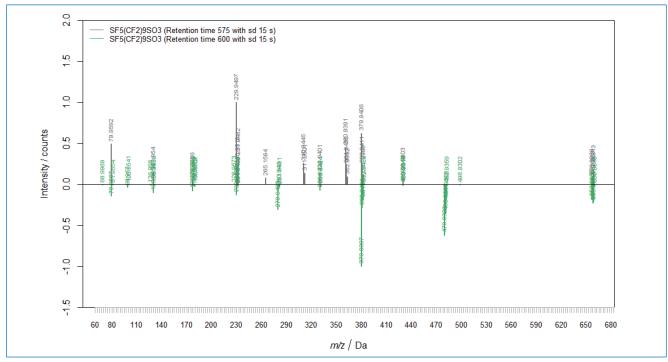


Figure 6: Comparison of the averaged fragmentation patterns of the ion trace  $SF_5(CF_2)_9SO_3^-$  at 575 +/-15 seconds (grey) and at 600 +/-15 seconds (green).

Table 5: Identifiable fragments of the averaged fragmentation patterns of the ion trace  $SF_5(CF_2)_gSO_3^-$  at 575 and 600 seconds.

575 s		600 s		
Fragment	m/z	Fragment	m/z	
SF <sub>5</sub>	79.9592 Da	(CF <sub>2</sub> ) <sub>4</sub> SO <sub>3</sub> -	279.9440 Da	
(CF <sub>2</sub> ) <sub>3</sub> SO <sub>3</sub> -	229.9472 Da	$(CF_2)_6SO_3^{-1}$	379.9377 Da	
(CF <sub>2</sub> ) <sub>6</sub> SO <sub>3</sub> -	379.9377 Da	(CF <sub>2</sub> ) <sub>8</sub> SO <sub>3</sub> -	479.9313 Da	

### **Conclusion**

The YMC-Triart C18 column demonstrates robust performance for PFAS analysis. It delivers:

- Efficient separation of key PFAS standards with linear quantification.
- Reproducible results in complex matrices, such as soil extracts.
- Improved isomer separation, enabling structural elucidation through distinct fragmentation patterns.

The column provides a reliable and enhanced approach to targeted and non-targeted PFAS analysis, supporting environmental monitoring and regulatory compliance.

#### References:

[1] Environ. Sci. Technol. 2023, 57, 6647-6655.

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<sup>\*</sup> Application data by courtesy of Ricardo Cunha, Institut für Umwelt & Energie, Technik & Analytik e. V. (IUTA) Duisburg, Germany, Boris Bugsel and Jonathan Zweigle.